Supplementary Table 1. Characterization of *Wolbachia* infection across beetle species and populations based on data in the literature.

Citation	Examined- infected species	taxonomy	Geography	*Wolbachia* genotyping	host genotyping	number of examined populations	number of examined individuals	species level	population level	individuals level	supergroup	effects fon hosts	other effects
Kageyama et al. 2010	*Lasioderma serricorne*	Anobiidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2015	*Lasioderma serricorne*	Anobiidae	Canada,Europe,USA	wsp,MLST	n.n.	5	n.n.	all infected	n.n.	n.n.	n.n.	n.n.	n.n.
Kageyama et al. 2010	*Stegobium paniceum*	Anobiidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2015	*Stegobium paniceum*	Anobiidae	Canada,USA	wsp,MLST	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Sontowski et al. 2015	*Agrilus araxenus Iablokoff*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	F	n.n.	n.n.
Sontowski et al. 2015	*Agrilus ater*	Buprestidae	France	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Agrilus cuprescens*	Buprestidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Agrilus derasofasciatus*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Agrilus graminis*	Buprestidae	France	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Agrilus ribesi*	Buprestidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Anthaxia anatolica*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Capnodis tenebricosa*	Buprestidae	Armenia, France	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Chrysobothris affinis*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Lamprodila mirifica nadezhdae*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Sphaerobothris aghababiani*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	F	n.n.	n.n.
Sontowski et al. 2015	*Sphenoptera antiqua*	Buprestidae	Armenia	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Trachys minutus*	Buprestidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Malloch et al. 2000	*Byturus tomentosus*	Byturidae	Europe	ftsZ	n.n.	34	152	n.n.	24% or 99% infected	97% multiple infected	A	n.n.	n.n.
Heintzman et al. 2014	*Amara alpina*	Carabidae	Arctica	Illumina shot-gun seq	n.n.	n.n.	6	n.n.	n.n.	2 strains	AB	n.n.	n.n.
Frank et al. 2009	*Pheropsophus aequinoctialis*	Carabidae	S America	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Aikawa et al. 2009	*Monochamus alternatus*	Cerambycidae	Japan	ftsZ,wsp,16S,groEL	n.n.	2	25	4% population infected	22/25 infected	single ftsZ variant	A	14% of W. genes transferred into autosomes of the host - no living bacteria detected	n.n.
Clark et al. 2001	*Acalymma blandulum*	Chrysomelidae	N America	16S, ftsZ	n.n.	1	20	all infected	n.n.	n.n.	A	n.n.	n.n.
Clark et al. 2001	*Acalymna vittatum*	Chrysomelidae	N America	16S, ftsZ	n.n.	1	20	5% infected	n.n.	n.n.	A	n.n.	n.n.
Werren et al. 1995	*Acronis sparsa*	Chrysomelidae	Panama	ftsZ	28S	n.n.	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Pankewitz et al. 2007	*Agelastica alni*	Chrysomelidae	Germany	16S,wsp,ftsZ	n.n.	1	5	all infected	all infected	n.n.	A	n.n.	n.n.
Jäckel et al. 2013	*Altica aenescens*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	3	n.n.	single strain (3/3 infected)	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica birmanensis*	Chrysomelidae	Taiwan	ftsZ	COI, ITS2, EF1α, msαts	1	1	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica brevicollis*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	1	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica carinata*	Chrysomelidae	USA	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica carinthiaca*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Xue et al. 2011	*Altica cirsicola*	Chrysomelidae	East Asia	wsp	COI,ITS2,EF1α	2	23	all infected	all infected	single infected	n.n.	n.n.	n.n.
Jäckel et al. 2013	*Altica engstroemi*	Chrysomelidae	Sweden	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	single strain	single infected	B	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica ericeti*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Xue et al. 2011	*Altica fragariae*	Chrysomelidae	East Asia	wsp	COI,ITS2,EF1α	4	47	all infected	all infected	single infected	n.n.	n.n.	n.n.
Jäckel et al. 2013	*Altica helianthemi*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	1	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica impressicollis*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	single strain (5/9 infected)	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica lythri*	Chrysomelidae	Germany; Poland; France	ftsZ	COI, ITS2, EF1α, msαts	18	50	n.n.	single strain (19/50 infected)	single infected or uninfected females	AB	perfect LD between W. infection and mtDNA haplotype (SS), CI, sex ratio distortion	horizontal transmission?
Jäckel et al. 2013	*Altica oleracea*	Chrysomelidae	Europe	ftsZ	COI, ITS2, EF1α, msαts	4	24	n.n.	single strain (10/24 infected)	single infected or uninfected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica palustris*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	9	n.n.	single strain	single infected	A	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica quercetorum*	Chrysomelidae	Germany	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	two strains	double infected	AB	n.n.	horizontal transmission?
Jäckel et al. 2013	*Altica tamaricis*	Chrysomelidae	France	ftsZ	COI, ITS2, EF1α, msαts	1	2	n.n.	single strain	single infected	n.n.	n.n.	horizontal transmission?
Xue et al. 2011	*Altica viridicyanea*	Chrysomelidae	East Asia	wsp	COI,ITS2,EF1α	3	57	all infected	all infected	single infected	n.n.	n.n.	n.n.
Roehrdanz et al. 2006	*Aphthona nigriscutis*	Chrysomelidae	USA, Canada	wsp,ftsZ	16S	3	214	c. 70% infected	60%-94% infected in populations	single strain	A	SS and reduction of host diversity	n.n.
Takano et al. 2017	*Brontispa longissima*	Chrysomelidae	East Timor	16S (Illumina),MLST	n.n.	2	12	n.n.	n.n.	n.n.	n.n.	CI	identification of new Alphaproteobacteria
Kondo et al. 2011	*Callosobruchus analis*	Chrysomelidae	Britain, Sri Lanka, Bangladesh	wsp,ftsZ	mtDNα	3	8	all infected	n.n.	n.n.	B	n.n.	n.n.
Kageyama et al. 2010	*Callosobruchus analis*	Chrysomelidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Numajiri et al. 2017	*Callosobruchus analis*	Chrysomelidae	Sri Lanca	wsp	COI	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	CI, fitness decline in infected beetles	n.n.
Kageyama et al. 2010	*Callosobruchus chinensis*	Chrysomelidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Nikoh et al. 2008	*Callosobruchus chinensis*	Chrysomelidae	Japan, Taiwan	wsp,ftsZ	n.n.	8 strains	14	n.n.	n.n.	n.n.	n.n.	30% of W. genes ransferred into autosomes of the host	n.n.
Kondo et al. 2011	*Callosobruchus chinensis*	Chrysomelidae	Asia, Africa	wsp,ftsZ	mtDNα	37	184	all infected	n.n.	n.n.	AB	n.n.	n.n.
Kondo et al. 1999	*Callosobruchus chinensis*	Chrysomelidae	Japan	16S, wsp, ftsZ	mtDNα	6	409	all infected	all infected		n.n.	n.n.	n.n.
Kondo et al. 2002	*Callosobruchus chinensis*	Chrysomelidae	Japan	wsp,ftsZ	n.n.	9	622	all infected	all infected	3 strains, infected: 93.7% triple , 6.1% double, 0.2% single, 1 strain in 90-100% ind.	AB	CI, W. populations (strains) controled by the host	n.n.
Kondo et al. 2005	*Callosobruchus chinensis*	Chrysomelidae	Japan	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Okayama et al. 2016	*Callosobruchus chinensis*	Chrysomelidae	Japan	n.n.	n.n.	1 (single strain)	n.n.	n.n.	n.n.	n.n.	n.n.	infection affects the life history and sexual selection (CI)	n.n.
Goodacre et al. 2015	*Callosobruchus rhodesianus*	Chrysomelidae	Africa,Asia, Australia,N America	wsp	COI	1	4	n.n.	1/4 infected	n.n.	n.n.	n.n.	n.n.
Kondo et al. 2011	*Callosobruchus latealbus*	Chrysomelidae	China, Taiwan	wsp,ftsZ	mtDNα	3	11	all infected	n.n.	2 strains, 10 ind. single infected, 2 ind. double infected	AB	n.n.	n.n.
Werren et al. 2000	*Charidotella purpurata*	Chrysomelidae	USA	ftsZ,16S	n.n.	n.n.	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Werren et al. 1995	*Chelymorpha alternans*	Chrysomelidae	Panama	ftsZ	28S	n.n.	3	n.n.	n.n.	n.n.	B	n.n.	n.n.
Keller et al. 2004	*Chelymorpha alternans*	Chrysomelidae	Panama	16S, wsp, ftsZ	COI	24	753	747/753 infected	infected by two strains	99.2% and 53% of beetles infected by 2 strains, single or double infected	n.n.	CI	dry climate decrease infection frequency
Clark et al. 2008	*Chelymorpha alternans*	Chrysomelidae	Panama	16S, wsp	n.n.	2	n.n.	n.n.	n.n.	n.n.	n.n.	W. modification of sperm	n.n.
Werren et al. 1995	*Chersinelina heteropunctata*	Chrysomelidae	Panama	ftsZ	28S	n.n.	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Kubisz et al. 2012	*Crioceris quaterdecimpunctata*	Chrysomelidae	C,E Europe	ftsZ,hcpA	COI, ITS1	5	25	all infected	all infected	most double infected	AB	n.n.	n.n.
Kolasa et al. 2017	*Crioceris quaterdecimpunctata*	Chrysomelidae	C,E Europe	MLST	COI,ITS1	4	19	all infected	all infected	all double infected	AB	n.n.	host plant transmission (Asparagus)
Kolasa et al. 2017	*Crioceris quinquepunctata*	Chrysomelidae	C,E Europe	MLST	COI,ITS1	2	10	all infected	all infected	all double infected	AB	n.n.	host plant transmission (Asparagus)
Mazur et al. 2014	*Crioceris quinquepunctata*	Chrysomelidae	C,E Europe	ftsZ,hcpA	COI,ITS1	2	10	all infected	all infected	most double infected	AB	n.n.	n.n.
Roehrdanz & Levine 2007	*Diabrotica barberi*	Chrysomelidae	USA	16S, wsp, ftsZ	mtDNα	66	577	all infected	all infected	2 distinct strains	A	CI	n.n.
Roehrdan & Wichmann 2013	*Diabrotica barberi*	Chrysomelidae	USA	wsp	n.n.	66	577	all infected	all infected	5 wsp variants	n.n.	n.n.	n.n.
Clark et al. 2001	*Diabrotica cristata*	Chrysomelidae	N America	16S,ftsZ	n.n.	1	25	12% infected	n.n.	n.n.	A	n.n.	n.n.
Clark et al. 2001	*Diabrotica lemniscata*	Chrysomelidae	N America	16S,ftsZ	n.n.	1	20	all infected	n.n.	n.n.	A	n.n.	n.n.
Clark et al. 2001	*Diabrotica virgifera virgifera*	Chrysomelidae	N America	16S,ftsZ	n.n.	1	40	all infected	n.n.	n.n.	A	possible CI	n.n.
Giordano et al. 1997	*Diabrotica virgifera virgifera*	Chrysomelidae	N America	16S,ftsZ	n.n.	15	60	13/15 populations infected	n.n.	n.n.	A	CI	n.n.
Barr et al. 2010	*Diabrotica virgifera virgifera*	Chrysomelidae	N America	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	W. down-regulate defence genes in maize
Pankewitz et al. 2007	*Galeruca tanaceti*	Chrysomelidae	Germany	16S,wsp,ftsZ	n.n.	1	5	all infected	all infected	n.n.	A	n.n.	n.n.
Jäckel et al. 2013	*Hermaeophaga mercurialis*	Chrysomelidae	n.n.	ftsZ	COI,ITS2,EF1α	1	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Kondo et al. 2011	*Megabruchidius sophorae*	Chrysomelidae	Japan	wsp,ftsZ	mtDNA	1	8	1/8 infected	n.n.	n.n.	B	n.n.	n.n.
Chafee et al. 2009	*Neochlamisus bebbianae*	Chrysomelidae	N America	MLST	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Montagna et al. 2014	*Oreina bidentata*	Chrysomelidae	Italy	16S,wsp,MLST	COI	1	1	all infected	all infected	single strain	A	n.n.	n.n.
Montagna et al. 2014	*Oreina cacaliae*	Chrysomelidae	Italy	16S,wsp,MLST	COI	1	15	all infected	all infected	single strain	A	n.n.	n.n.
Montagna et al. 2014	*Oreina elongate*	Chrysomelidae	Italy	16S,wsp,MLST	COI	2	3	all infected	all infected	single strain	A	n.n.	n.n.
Montagna et al. 2014	*Oreina liturata*	Chrysomelidae	Italy	16S,wsp,MLST	COI	1	1	all infected	all infected	single strain	A	n.n.	n.n.
Montagna et al. 2014	*Oreina speciosa*	Chrysomelidae	Italy	16S,wsp,MLST	COI	3	5	all infected	all infected	single strain	A	n.n.	n.n.
Jäckel et al. 2013	*Podagrica fuscipes*	Chrysomelidae	n.n.	ftsZ	COI,ITS2,EF1α	1	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Yun et al. 2011	*Propylea japonica*	Chrysomelidae	China	wsp	n.n.	n.n.	44	n.n.	n.n.	n.n.	B	n.n.	n.n.
Werren et al. 1995	*Priocera sp.*	Cleridae	Panama	ftsZ	28S	n.n.	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Majerus et al. 2000	*Adalia bipunctata*	Coccinellidae	Russia	wsp	n.n.	1	110 (40 lines)	2 lines infected	n.n.	2 strains	B	killing of male embryo	coexistence with Rickettsia & Spiroplasma
Hurst et al. 1999	*Adalia bipunctata*	Coccinellidae	Russia	ftsZ,wsp,16S	n.n.	1	5	n.n.	all infected	n.n.	B	n.n.	n.n.
Elnagdy et al. 2013	*Adalia bipunctata*	Coccinellidae	Russia	n.n.	n.n.	3 lines	40	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Weinert et al. 2007	*Calvia quattuordecimguttata*	Coccinellidae	Germany	wsp	n.n.	n.n.	>20	n.n.	4% of males infected	n.n.	n.n.	n.n.	coexistence with Rickettsia
Weinert et al. 2007	*Chilocorus bipustulatus*	Coccinellidae	Italy	wsp	n.n.	n.n.	>20	n.n.	8% of females infected	n.n.	n.n.	n.n.	coexistence with Spiroplasma
Weinert et al. 2007	*Coccidula rufa*	Coccinellidae	Germany	wsp	n.n.	n.n.	>20	n.n.	78% of females & 8% of males infected	n.n.	n.n.	n.n.	coexistence with Rickettsia
Weinert et al. 2007	*Coccinella septempunctata*	Coccinellidae	Britain	wsp	n.n.	n.n.	>20	n.n.	5% of females infected	n.n.	n.n.	n.n.	coexistence with Rickettsia
Elnagdy et al. 2013	*Coccinella undecimpunctata*	Coccinellidae	Egypt	n.n.	n.n.	6 lines	40	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Weinert et al. 2007	*Halyzia sedecimguttata*	Coccinellidae	Germany	wsp	n.n.	n.n.	>20	n.n.	2% of females infected	n.n.	n.n.	n.n.	coexistence with Rickettsia
Dudek et al. 2017	*Harmonia axyridis*	Coccinellidae	Poland	16S NGS (Illumina)	n.n.	1	15	n.n.	1/15 specimens infected	n.n.	n.n.	n.n.	coexistence with Rickettsia & Spiroplasma
Goryachevaa et al. 2015	*Harmonia axyridis*	Coccinellidae	Europe, Japan, USA	MSLT, wsp	n.n.	13	375	4 out of 13 infected	n.n.	n.n.	AB	n.n.	n.n.
Weinert et al. 2007	*Rhizobius litura*	Coccinellidae	Germany	wsp	n.n.	n.n.	>20	n.n.	89% of females & 5% of males infected	n.n.	n.n.	n.n.	coexistence with Rickettsia
Rodriguer et al. 2010a	*Aramigus conirostris*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single strain	B	only parthenogenetic or presumed parthenogenetic species infected	n.n.
Rodriguer et al. 2010a	*Aramigus tessellatus*	Curculionidae	S America	MLST	COI	n.n.	2	all infected	all infected	single strain	B	n.n.	n.n.
Toju et al. 2013	*Archarius roelofsi*	Curculionidae	Asia	16S	n.n.	3	3	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Rodriguer et al. 2010a	*Atrichonotus taeniatulus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single strain	B	n.n.	n.n.
Campbell et al. 1992	*Bangasternus orientalis*	Curculionidae	Italy	16S	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Barypeithes pellucidus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Brachyderes incanus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Piper et al. 2001	*Cathormiocerus britannicus*	Curculionidae	England	16S	ITS2	3	5	all infected	all infected	n.n.	n.n.	n.n.	n.n.
Piper et al. 2001	*Cathormiocerus myrmecophilus*	Curculionidae	England	16S	ITS2	3	3	all infected	all infected	n.n.	n.n.	n.n.	n.n.
Floate et al. 2011	*Ceutorhynchus obstrictus*	Curculionidae	Europe, N America	MSLT,wsp	COI	11	64	all infected	all infected	n.n.	n.n.	n.n.	n.n.
Zhang et al. 2010	*Conotrachelus nenuphar*	Curculionidae	USA	wsp,ftsZ	COI	12	94	98% infected	n.n.	3 strains (double and single infected)	AB	possible CI	n.n.
Werren et al. 1995	*Cossomus sp.*	Curculionidae	Panama	ftsZ	28S	n.n.	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Zchori-Fein et al. 2006	*Coccotrypes dactyliperda*	Curculionidae	Israel	16S	n.n.	1	20	n.n.	all infected	n.n.	n.n.	role in oogenesis	n.n.
Toju et al. 2013	*Curculio hachijoensis*	Curculionidae	Asia	16S	n.n.	1	2	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Toju et al. 2013	*Curculio hilgendorfi*	Curculionidae	Asia	16S	n.n.	2	3	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Toju et al. 2013	*Curculio morimotoi*	Curculionidae	Asia	16S	n.n.	1	2	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.

Toju et al. 2013	*Curculio okumai*	Curculionidae	Asia	16S	n.n.	2	4	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Kajtoch et al. 2017	*Cyanapion afer*	Curculionidae	Poland	MLST	COI, ITS2, EF1α	5	10	all infected	all infected	2 strains	A	n.n.	n.n.
Kajtoch et al. 2017	*Cyanapion gyllenhalii*	Curculionidae	Poland	MLST	COI, ITS2, EF1α	6	12	all infected	all infected	single strain	A	n.n.	n.n.
Kajtoch et al. 2017	*Cyanapion spencii*	Curculionidae	Poland	MLST	COI, ITS2, EF1α	6	12	n.n.	5/6 infected	2 strains	AB	n.n.	n.n.
Werren et al. 2000	*Cyrtepistomus castaneus*	Curculionidae	USA	ftsZ,16S	n.n.	n.n.	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Dorytomus carpathicus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Dorytomus rufatus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Ellescus bipunctatus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Rodriguer et al. 2010a	*Eurymetopus fallax*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single strain	B	n.n.	n.n.
Rodriguer et al. 2010a	*Eurymetopus globosus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single strain	B	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Eusomus ovulum*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	-	n.n.	A	n.n.	n.n.
Mazur et al. 2016	*Eusomus ovulum*	Curculionidae	C,SE,E Europe	MLST	COI, ITS2, EF1α	28	65	all infected	all infected, single strain	all infected, single strain	A	SS, possible pathernogenesis induction	n.n.
Kawasaki et al. 2016	*Euwallacea interjectus*	Curculionidae	Japan	wsp,MLST	COI, EF1α	n.n.	20	n.n.	100% infected	single infected	A	haplodiploidy induction uncertain	horizontal transmission?
Kawasaki et al. 2016	*Euwallacea validus*	Curculionidae	Japan	wsp,MLST	COI, EF1α	n.n.	21	n.n.	75% infected	single infected	A	haplodiploidy induction uncertain	horizontal transmission?
Berasategui et al. 2016	*Hylobius abietis*	Curculionidae	Europe	16S NGS(454)	n.n.	6	54	all infected	0.2-100% abundance, 28-73% of reads	n.n.	n.n.	n.n.	n.n.
O'Neill et al. 1992	*Hypera postica*	Curculionidae	n.n.	16S	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Iwase et al.. 2015	*Hypera postica*	Curculionidae	Japane, Europe	wsp,ftsZ,coxA,hpcA	CytB	11	242	5/11 populations infected	0-100% infected	2 strains	B	n.n.	n.n.
Mariño et al. 2017	*Hypothenemus hampei*	Curculionidae	Puerto Rico	wsp	n.n.	1	n.n.	n.n.	n.n.	n.n.	n.n.	CI	W. as biocontrol agent
Vega et al. 2002	*Hypothenemus hampei*	Curculionidae	S & C America, Asia, Africa	wsp	n.n.	17 countries	n.n.	n.n.	11/17 populations infected	possibly only single strain	B	possible sex determination	n.n.
Toju et al. 2013	*Koreoculio minutissimus*	Curculionidae	Asia	16S	n.n.	2	3	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Chen et al. 2012	*Lissorhoptrus oryzophilus*	Curculionidae	China, Texas	wsp,MLST	n.n.	6	127	all infected	all infected	all infected	B	W. necessary for oocyte production	n.n.
Lu et al. 2014	*Lissorhoptrus oryzophilus*	Curculionidae	China, Texas	16S	n.n.	5	1421	all infected	all infected	99 infected	n.n.	n.n.	n.n.
Rodriguer et al. 2010a	*Mimographus ocellatus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Prakash & Puttaraju 2006	*Myllocerus discolor*	Curculionidae	India	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus ambiguus*	Curculionidae	S America	MLST	COI	n.n.	3	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus cervinus*	Curculionidae	S America	MLST	COI	n.n.	20	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguero et al. 2010b	*Naupactus cervinus*	Curculionidae	S America	16S, wsp, coxA, fbpA	COI, ITS1n.n.18S	39	309	all infected	all infected	single infected	B	LD	host bottleneck caused by W.
Rodriguer et al. 2010a	*Naupactus condecoratus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus cyphoides*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus dissimilis*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus leucoloma*	Curculionidae	S America	MLST	COI	n.n.	3	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus minor*	Curculionidae	S America	MLST	COI	n.n.	3	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus peregrinus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus purpureoviolaceus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus tremolerasi*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Naupactus verecundus*	Curculionidae	S America	MLST	COI	n.n.	1	all infected	all infected	single infected	B	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Otiorhynchus coecus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Stenberg & Lundmark 2004	*Otiorhynchus scaber*	Curculionidae	Austria, Slovenia	16S	COI,COII,CytB	2	28	all infected	infected: 5/10 ♂ , 2/7 sexual ♀, 0 of clonal ♀	n.n.	B	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Otiorhynchus singularis*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Son et al. 2008	*Otiorhynchus sulcatus*	Curculionidae	USA	wsp	n.n.	5	96	all infected	all infected	n.n.	B	W. necessary for egg developnment	n.n.
Rodriguer et al. 2010a	*Pantomorus auripes*	Curculionidae	S America	MLST	COI	n.n.	3	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Pantomorus cinerosus*	Curculionidae	S America	MLST	COI	n.n.	2	all infected	all infected	single infected	B	n.n.	n.n.
Rodriguer et al. 2010a	*Pantomorus viridisquamosus*	Curculionidae	S America	MLST	COI	n.n.	2	all infected	all infected	single infected	B	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Paophilus aflatus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Parafoucartia squamulata*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Arthofer et al. 2009	*Pityogenes chalcographus*	Curculionidae	C,E,N,S Europe	wsp, MLST	COI, EF1α	31	344	35.5% infected	0-100% infected	3 strains (91% - A, 9% - B)(95% single infected, 5% double infected)	AB	n.n.	bacteria loss and horizontal transfer
Lachowska-Cierlik et al. 2010	*Polydrusus inustus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	A	n.n.	n.n.
Kajtoch et al. 2012	*Polydrusus inustus*	Curculionidae	C,E Europe	wsp,16S,ftsZ,hcpA	COI, ITS2,EF1α	9	40	all infected	all infected, 2 closely related strains	all infected, 2 closely related strains	A	probably SS	n.n.
Lachowska-Cierlik et al. 2010	*Polydrusus mollis*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Kajtoch et al. 2012	*Polydrusus pilifer*	Curculionidae	Caucasus	wsp,16S,ftsZ,hcpA	COI, ITS2,EF1α	1	5	all infected	all infected	single infected	A	probably SS	n.n.
Lachowska-Cierlik et al. 2010	*Polydrusus pilosus*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	AB	n.n.	n.n.
Campbell et al. 1992	*Rhinocyllus conicus*	Curculionidae	Italy	16S	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	F	n.n.	n.n.
Toševski et al. 2015	*Rhinusa pilosa*	Curculionidae	Europe	coxA	COI, 16S,EF1α	6	47	n.n.	80.8% ind. Infected	n.n.	n.n.	n.n.	n.n.
Toševski et al. 2015	*Rhinusa rara*	Curculionidae	Europe	coxA	COI, 16S,EF1α	5	10	n.n.	60% ind. Infected	n.n.	n.n.	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Sciaphobus rubi*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
White et al. 2015	*Sitona obsoletus*	Curculionidae	New Zealand	16S(454),MLST	n.n.	2	36	all infected	all infected	W. = 31-72% of bacteria reads	A	n.n.	coexistence with Rickettsia
Heddi et al. 1999	*Sitophilus granarius*	Curculionidae	France	16S	n.n.	3	30	2 out of 3 infected	40% infected	n.n.	n.n.	n.n.	n.n.
Heddi et al. 1999	*Sitophilus oryzae*	Curculionidae	Africa,Asia, Australia,N America	16S	n.n.	10	100	7 out of 10 infected	61% infected	n.n.	B	n.n.	n.n.
Carvalho et al. 2014	*Sitophilus oryzae*	Curculionidae	America S, N, Asia, Australia	16S	16S	16	n.n.	n.n.	7/16 infected	n.n.	n.n.	n.n.	horizontal transmission
Li et al. 2015	*Sitophilus oryzae*	Curculionidae	Canada	wsp,MLST	n.n.	7	n.n.	1/6 populations infected	n.n.	n.n.	n.n.	n.n.	n.n.
Werren et al. 1995	*Sitophilus oryzae*	Curculionidae	Panama	ftsZ	28S	n.n.	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Carvalho et al. 2014	*Sitophilus zaemais*	Curculionidae	America S, N, Asia, Australia	16S	16S	11	n.n.	n.n.	9/11 infected	n.n.	n.n.	n.n.	horizontal transmission
Kageyama et al. 2010	*Sitophilus zaemais*	Curculionidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2015	*Sitophilus zaemais*	Curculionidae	Thailand	wsp,MLST	n.n.	3	n.n.	2/3 populations infected	n.n.	n.n.	n.n.	n.n.	n.n.
Heddi et al. 1999	*Sitophilus zaemais*	Curculionidae	Asia, Africa, America,	16S	n.n.	10	100	4 out of 10 infected	40% infected	n.n.	n.n.	n.n.	n.n.
White et al. 2015	*Steriphus variabilis*	Curculionidae	New Zealand	16S(454),MLST	n.n.	1	18	all infected	all infected	W. = 8-12% of bacteria reads	B	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Strophosoma capitatum*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Lachowska-Cierlik et al. 2010	*Strophosoma melanogrammum*	Curculionidae	Poland	wsp,16S,ftsZ,hcpA	COI	1	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Kawasaki et al. 2016	*Taphrorychus bicolor*	Curculionidae	Austria	wsp,MLST	COI,EF1α	n.n.	5	n.n.	80% infected	single infected	B	n.n.	horizontal transmission?
Kawasaki et al. 2016	*Xyleborinus schaufussi*	Curculionidae	Japan	wsp,MLST	COI,EF1α	n.n.	11	n.n.	85% infected	single infected	A	haplodiploidy induction uncertain	horizontal transmission?
Kawasaki et al. 2016	*Xyleborus dispar*	Curculionidae	Austria	wsp,MLST	COI,EF1α	n.n.	5	n.n.	100% infected	6 strains, ind. multiple infected	A	haplodiploidy induction uncertain	horizontal transmission?
Kawasaki et al. 2016	*Xyleborus seiryorensis*	Curculionidae	Japan	wsp,MLST	COI,EF1α	n.n.	7	n.n.	100% infected	single infected	A	haplodiploidy induction uncertain	horizontal transmission?
Kawasaki et al. 2016	*Xylosandrus crassiusculus*	Curculionidae	Japan	wsp,MLST	COI,EF1α	n.n.	47	n.n.	95% infected	single infected	A	haplodiploidy induction uncertain	horizontal transmission?
Kawasaki et al. 2016	*Xylosandrus germanus*	Curculionidae	Japane	wsp,MLST	COI	9	120	all infected	all infected, 5 strains, 1-4 strains/population	9 allele combinations	A	multi-directional CI	n.n.
Kawasaki et al. 2016	*Xylosandrus germanus*	Curculionidae	Germany, Hungary	wsp,MLST	COI,EF1α	n.n.	130	n.n.	100% infected	4 strains, ind. multiple infected	A	haplodiploidy induction uncertain	horizontal transmission?
Kageyama et al. 2010	*Anthrenus verbasci*	Dermestidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2015	*Attagenus unicolor*	Dermestidae	USA	wsp,MLST	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	
Li et al. 2015	*Dermestes lardarius*	Dermestidae	Canada	wsp,MLST	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	
García-Vázquez & Ribera 2016	*Deronectes angelinii*	Dytiscidae	Italy	wsp	COI,16S,NADH,H3,Wg	1	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
García-Vázquez & Ribera 2016	*Deronectes aubei*	Dytiscidae	Italy, Germany, Spain, France	wsp	COI,16S,NADH,H3,Wg	5	15	8/15 infected	n.n.	n.n.	A	n.n.	n.n.
García-Vázquez & Ribera 2016	*Deronectes delarouzei*	Dytiscidae	Spain	wsp	COI,16S,NADH,H3,Wg	4	4	all infected	n.n.	n.n.	A	n.n.	n.n.
García-Vázquez & Ribera 2016	*Deronectes moestus*	Dytiscidae	Greece, Italy, Spain, Marocco	wsp	COI,16S,NADH,H3,Wg	21	21	11/21 infected	n.n.	n.n.	A,B	n.n.	n.n.
García-Vázquez & Ribera 2016	*Deronectes semirufus*	Dytiscidae	Italy	wsp	COI,16S,NADH,H3,Wg	5	6	all infected	n.n.	n.n.	A,B	n.n.	n.n.
Sontowski et al. 2015	*Agabus bipustulatus*	Dytiscidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Copelatus haemorrhoidalis*	Dytiscidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hygrotus versicolor*	Dytiscidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Gyrinus suffriani*	Gyrinidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Haliplus fluviatilis*	Haliplidae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena brachymera*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena corinna*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena gracilis*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena iberica*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena inapicipalpis*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena morio*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena pygmaea*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Hydraena riberai*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Limnebius atomus*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Limnebius truncatellus*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Ochthebius exculptus*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Ochthebius meridionales*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Ochthebius minimus*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	B	n.n.	n.n.
Sontowski et al. 2015	*Ochthebius tivelunus*	Hydraenidae	n.n.	MLST	18S,28S,COI	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Sontowski et al. 2015	*Berosus luridus*	Hydrophilidae	n.n.	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Jeong et al. 2009	*Luciola unmunsana*	Lampyridae	Korea	16S,wsp	n.n.	5	10	n.n.	80% infected	n.n.	A	n.n.	n.n.
Werren et al. 2000	*Epicauta cinerea*	Meloidae	USA	ftsZ,16S	n.n.	n.n.	1	n.n.	n.n.	n.n.	B	n.n.	n.n.
Perotti et al. 2016	*Micromalthus debilis*	Micromalthidae	USA	16S,wsp	n.n.	1	62	n.n.	3/62 infected	n.n.	n.n.	parthenogenetic paedogenesis induction?	coexistence with Rickettsia
Sontowski et al. 2015	*Noterus clavicornis*	Noteridae	Germany	ftsZ	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	A	n.n.	n.n.
Jensen 2011	*Popillia japonica*	Scarabaeidae	Japane	wsp	COI	5 plant hosts	182	n.n.	11.7% infected	n.n.	n.n.	possible CI	n.n.
Yun et al. 2011	*Paederus fuscipes*	Staphylinidae	China	wsp	n.n.	n.n.	7	n.n.	n.n.	n.n.	B	n.n.	n.n.
Bili et al. 2016	*Aleochara bilineata*	Staphylinidae	France	16S NGS(454), fbpA	n.n.	2	40	n.n.	n.n.	n.n.	A	n.n.	coexistence with Rickettsia & Spiroplasma
Bili et al. 2016	*Aleochara bipustulata*	Staphylinidae	France	16S NGS(454), fbpA	n.n.	2	40	n.n.	n.n.	n.n.	B	n.n.	coexistence with Spiroplasma
Li et al. 2015	*Oryzaephilus mercator*	Sylvanidae	Canada	wsp,MLST	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Sharaf et al. 2010	*Oryzaephilus surinamensis*	Sylvanidae	Israel	16S	n.n.	2	94	69-86% infected	n.n.	n.n.	n.n.	possible CI	n.n.
Sharaf et al. 2013	*Oryzaephilus surinamensis*	Sylvanidae	Israel	16S	COI,16S	3	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2015	*Oryzaephilus surinamensis*	Sylvanidae	Canada,Europe,USA	wsp,MLST	n.n.	5	n.n.	all infected	n.n.	n.n.	n.n.	n.n.	n.n.
Kageyama et al. 2010	*Oryzaephilus surinamensis*	Sylvanidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.

Kageyama et al. 2010	*Tribolium confusum*	Tenebrionidae	Japane	wsp	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Goodacre et al. 2015	*Tribolium confusum*	Tenebrionidae	USA, China, Uganda	wsp	COI	9	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2016	*Tribolium confusum*	Tenebrionidae	Canada	wsp	n.n.	colonies	80	all infected?	100% infected?	n.n.	n.n.	CI	n.n.
Ming et al. 2015	*Tribolium confusum*	Tenebrionidae	China	wsp	EF-2	1	numerous	n.n.	n.n.	n.n.	n.n.	CI	n.n.
Fialho & Stevens 1997	*Tribolium confusum*	Tenebrionidae	Asia, USA, Europe	ftsZ,ITS	n.n.	11	-	all infected	all infected	n.n.	B	n.n.	n.n.
Fialho &Stevens 1996	*Tribolium confusum*	Tenebrionidae	Asia, USA, Europe	16S	n.n.	8 strains	n.n.	n.n.	n.n.	n.n.	n.n.	CI	n.n.
O'Neill et al. 1992	*Tribolium confusum*	Tenebrionidae	n.n.	16S	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.	n.n.
Li et al. 2015	*Tribolium confusum*	Tenebrionidae	Canada,Europe,USA	wsp,MLST	n.n.	16	n.n.	15/16 populations infected	n.n.	n.n.	n.n.	n.n.	n.n.
Fialho & Stevens 2000	*Tribolium madens*	Tenebrionidae	n.n.	ftsZ,wsp,ITS,groEL	n.n.	3 strains	n.n.	2 strains infected	n.n.	n.n.	n.n.	male-killing	n.n.

Abbreviations: W. – *Wolbachia* , LD - linkage disequilibrium, SS- selective sweep, CI - cytoplasmic incompatibility, MLST – multilocus sequence typing, gatB - aspartyl/glutamyl-tRNA(Gln) amidotransferase, subunit B, coxA - cytochrome c oxidase, subunit I, hcpA - conserved hypothetical protein, ftsZ - cell division protein, fbpA - fructose-bisphosphate aldolase, wsp – *Wolbachia* surface protein, 16S/18S/28S rDNA - 16S/18S/28S ribosomal DNA, groEL - heat-shock protein 60, COI – cytochrome oxidase subunit I, ITS – internal transcribed spacer, EF1α – elongation factor 1α, Wg - wingless, H3 - histonr H3, msats – microsatellites.

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